ISOLATION OF SOME PHYTOCHEMICAL CONSTITUENTS FROM YACON TUBERS OF TAUNGGYI AREA

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Abstract

The *Smallanthus sonchifolius* (yacon) tubers were dried at the ambient temperature, chopped into bits and percolated with70% ethanol. Qualitative phytochemical screening was determined by the standard procedure suggested the presence of alkaloids, flavonoids, steroids, polyphenols, terpenoids, α -amino acids, glycosides, phenolic compounds, carbohydrate, saponins, tannins, protein, and reducing sugars. Starch and cyanogenic glycosides were not detected in yacon tubers. The profile of the chemical constituents present was established by thin layer chromatography. Six pure organic constituents namely, stigmasteryl acetate (0.003%), β -sitosteryl acetate (0.005%), hexadecanoic acid (0.009%), β -sitosterol (0.02%), chlorogenic acid (0.12%) and β -sitosterol β -D-glucoside (0.8%) based on ethyl acetate extract were isolated from the ethyl acetate soluble fraction of the 70% ethanol extract of the yacon tuber on a silica gel column by gradient elution with petroleum and ethyl acetate (PE-EA, 99:1 to EA only). The isolated compounds were characterized by UV and FTIR.

Keywords: Smallanthus sonchifolius, yacon tuber, percolation, phytochemical screening, ethyl acetate extract, UV, FTIR

Introduction

Yacon, *Smallanthus sonchifolius* (Poepp.&Endl.) H. Robinson, is a plant originally cultivated in South America, and the fresh root is eaten like a fruit in this area. Yacon was introduced to Japan in 1985 and has been gradually paid attention to due to its abundant content of fructooligo saccharide, which has some health-promoting effects such as improvement in the intestinal microflora balance, as a storage sugar in place of starch in its root

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(Nakanishi, 1997). Yacon tuber is sometimes used for home cooking but has not been a common foodstuff because of decaying easily or rapid browning of the juice or injured tissues. The browning may be caused by condensation reaction of polyphenols with amino compounds (Yabuta *et al.*, 2001) and enzymatic polymerization of polyphenols (Mayer, 1987). Yacon juice contains 850 ppm polyphenol compounds which generally have anti-oxidative activity and chlorogenic acid was reported as a major antioxidant in yacon as well as tryptophan (Yan *et al.*, 1999). In this study, investigation on ethyl acetate fraction of yacon tubers founds six compounds.

Materials and Methods

Smallanthus sonchifolius (yacon tuber) was collected from Naung Karr Village, Taunggyi Township, Southern Shan State in July 2016 and identified at Department of Botany, Taunggyi University. UV and IR spectra were recorded on Shimadzu 1800 UV-Vis Spectrometer and Shimadzu 8400 Fourier Transform- Infrared Spectrophotometer using KBr pellet. Aluminium backed silica gel GF₂₅₄precoated plates from Merck were used for TLC analysis.

Isolation of Compounds from YaconTubers

The yacon tuber was dried at ambient temperature and pulverized to coarse grained powder.Dry powder (120 g) was applied to extract with 70% ethanol using percolation method and the resultant crude extract obtained was filtered.The crude extract was concentrated under reduced pressure using a rotary evaporator to obtain a solvent free 70% ethanol extract. This extract (29 g) was stirred with ethyl acetate to obtain ethyl acetate soluble extract (0.5 g), which was separated on silica gel column by gradient elution with PE:EA (99:1, 9:1, 5:1, 3:1, 1:1, 1:2, 1:4 and EA only).Fractions obtained were collected in 10mL flasks. Identical fractions were pooled based on their TLC profiles thus resulting in 6 combined fractions.Further purification using methanol solvent was carried out on fractions that were impure.

Phytochemical Analysis

Qualitative phytochemical screening was carried out to determine the presence or absence of alkaloids, flavonoids, steroids, polyphenols, terpenoids, α -amino acids, glycosides, phenolic compounds, carbohydrate, saponins, tannins, starch, protein, reducing sugars, cyanogenic glycosides and using the standard literature procedure (William and Douglas, 2006).

Spectroscopic Analysis

The isolated compounds were characterized by Fourier transform infrared (FTIR) and ultraviolet spectroscopy. FTIR spectra were recorded in which samples were prepared as KBr pellets at the Universities' Research Centre, Yangon University. UV spectra were recorded on methanol solutions at the Chemistry Department, West Yangon University.

Results and Discussion

Phytochemical Analysis

The dried powder sample of yacon tuber showed the presence of various phytochemicals namely, alkaloids, flavonoids, steroids, polyphenols, terpenoids, α -amino acids, glycosides, phenolic compounds, carbohydrate, saponins, tannins, protein, and reducing sugars. Starch and cyanogenic glycosides were not detected in the sample (Table 1).

Table 1Phytochemicals Present in Tubers of Smallanthus
sonchifolius(Yacon)

| No. | Test | Extract | Reagent Used | Observation | Remarks |
|-----|-------------|---------|--|-------------|---------|
| 1 | Alkaloids | 1% HCl | Dragendorff's | Orange ppt. | + |
| | | | reagent | | |
| 2 | Flavonoids | 95% | Mg ribbon and | Pink color | + |
| | | EtOH | conc: H ₂ SO ₄ | | |
| 3 | Steroids | PE | Acetic anhydride | Blue color | + |
| | | | and conc: H ₂ SO ₄ | | |
| 4 | Polyphenols | EtOH | 1% FeCl ₃ and 1% | Greenish- | + |
| | | | $K_3[Fe(CN)_6]$ | blue color | |
| 5 | Terpenoids | EtOH | Acetic anhydride | Red color | + |
| | | | and conc: H ₂ SO ₄ | | |

| No. | Test | Extract | Reagent Used | Observation | Remarks |
|-----|--------------------------|---|--|-----------------------|---------|
| 6 | α- Amino acids | EtOH | Ninhydrin reagent | Violet color | + |
| 7 | Glycosides | H_2O | 10% lead acetate | White ppt. | + |
| 8 | Phenolic Compounds | H ₂ O | 10% FeCl ₃ | Brown color | + |
| 9 | Carbohydrates | H ₂ O | 10 % α-naphthol and conc: H_2SO_4 | Red ring | + |
| 10 | Saponins | H_2O | Distilled water | Frothing | + |
| 11 | Tannins | H ₂ O | 2% NaCl and 1% gelatin | Yellow brown color | + |
| 12 | Starch | H ₂ O | I ₂ solution | No blue color | — |
| 13 | Protein | Hot H ₂ O | NaOH and CuSO ₄ | Yellow ppt. | + |
| 14 | Reducing Sugars | H ₂ SO ₄ (dil) | NaOH and Benedict's | Brick red ppt. | + |
| 15 | Cyanogenic Glycosides | H ₂ O | solution conc: H ₂ SO ₄ , sodium picrate | No brick red | _ |
| | | | paper | | |

(+) = present, (-) = absent, (ppt.) = precipitate

Identification of the Isolated Compounds

After purification of the fractions from the silica gel column with methanol, six pure compounds were obtained. Thin layer chromatograms of six isolated compounds from EA extract of yacon tuber were identified by UV and IR spectroscopic methods. IR spectral database (AIST) was used for comparison.

The isolated compound $\underline{\mathbf{M}_{1}}$ could not be observed under short and long wavelength UV lights on TLC plate, but it gave a dark violet spot upon heating with anisaldehyde-sulphuric acid reagent (R_{f} 0.43, Silica gel GF₂₅₄, PE:EtOAc, 99:1). This suggests a steroid/terpenoid compound. The IR spectrum (Figure 1) of $\underline{\mathbf{M}_{1}}$ showed absorptions at 1739 and 1261 cm⁻¹ corresponding to C=O and C-O stretching vibrations of an acetate ester. Apart from this, the remaining bands are very similar to those of stigmasterol. From this observation, it may be deduced that $\underline{\mathbf{M}_{1}}$ is stigmasteryl acetate. The assignment of IR bands is summarized in Table (2). Therefore $\underline{\mathbf{M}_{1}}$ should be stigmasteryl acetate.

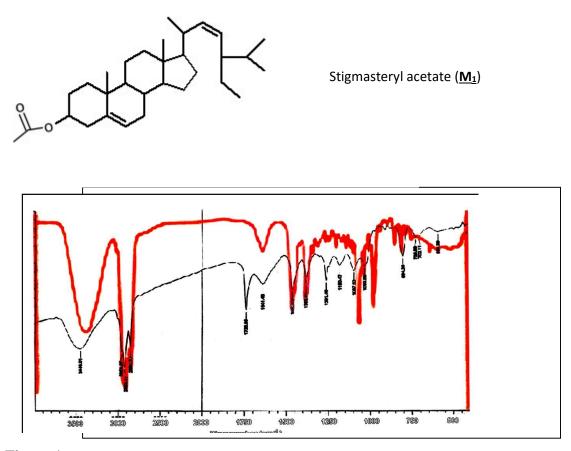


Figure 1 Overlaid IR spectra of M₁(black) and stigmasteryl acetate (red) (AIST)

The isolated compound $\underline{M_2}$ could not be observed under short and long wavelength UV lights on TLC plate, but it gave a dark violet spot upon heating with anisaldehyde-sulphuric acid reagent ($R_f 0.63$,Silica gel GF₂₅₄, PE:EtOAc, 99:1). This suggests a steroid/terpenoid compound. In the IR spectrum (Figure 2) of the compound, the O-H stretching band at 3447 cm⁻¹ of β -sitosterol disappeared, while new bands at 1738 cm⁻¹ for C = O stretching and 1287 cm⁻¹ for C-O stretching showed up, the remaining bands being similar to those of β -sitosterol. This can be seen in the overlaid IR spectra of $\underline{M_2}$ and β -sitosteryl acetate (Figure 2). The assignment of IR bands is summarized in Table (2). Therefore the isolated compound $\underline{M_2}$ should be β -sitosteryl acetate.

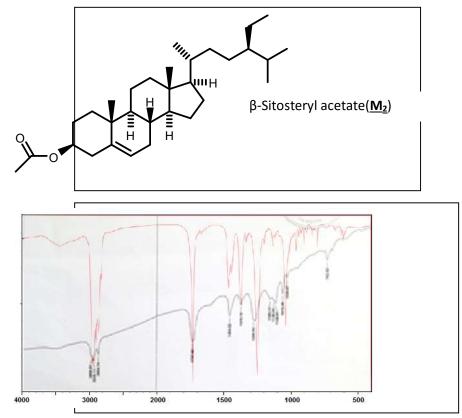


Figure 2 Overlaid IR spectra of M₂ (black) and β-sitosteryl acetate (red) (AIST)

The isolated compound \underline{M}_3 (R_f 0.65, Silica gel GF₂₅₄, PE:EtOAc, 9:1) was invisible under UV short and long wavelengths on TLC plate, but could be visualized as pink spot upon heating with anisaldehyde-sulphuric acid reagent. The IR spectrum of isolated compound \underline{M}_3 suggests a long chain carboxylic acid by the carbonyl stretching band, very broad OH stretching band and an intense CH₂ rocking band, respectively, at 3400-2400, 1697 and 723 cm⁻¹ (Figure 3). The assignment of bands is summarized in Table (3). From the close resemblance of the IR spectra (Figure 3) of \underline{M}_3 and hexadecanoic acid, \underline{M}_3 should be hexadecanoic acid. Hexadecanoic acid is also reported in the plant (Stuardxchange, 2016).

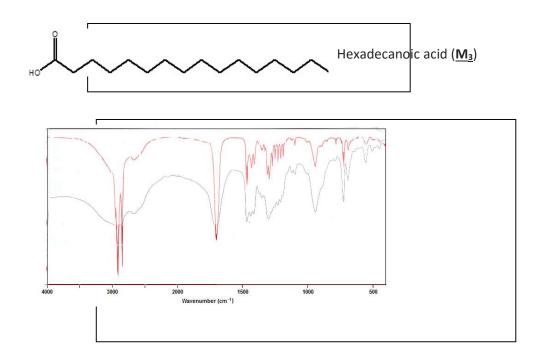


Figure 3 Overlaid IR spectra of M₃ (black) and hexadecanoic acid (red) (AIST)

The isolated compound $\underline{M}_4(R_f \ 0.44)$, Silica gel GF₂₅₄, PE:EtOAc, 5:1)was invisible under UV short and long wavelengths on TLC plate, but could be visualized as violet spot upon heating with anisaldehyde-sulphuric acid reagent, suggesting a steroid/terpenoid compound. The assignment of absorption bands in IR spectrum (Figure 4) is summarized in Table (2). From the close resemblance between the IR spectra of \underline{M}_4 and β -sitosterol (Figure 4), \underline{M}_4 should be β -sitosterol. This is also in accordance with a previous report on the plant (Stuardxchange, 2016).

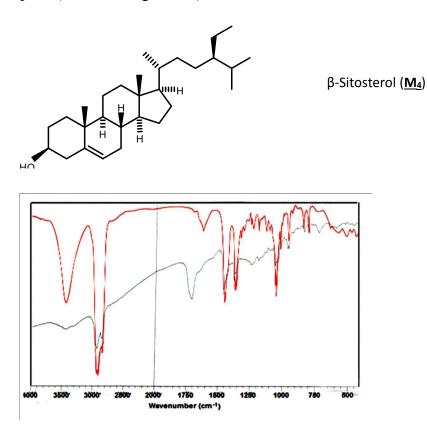


Figure 4 Overlaid IR spectra of M_4 (black) and β -sitosterol (red) (AIST)

The isolated compound $\underline{M_5}$ (R_f 0.42, Silica gel GF₂₅₄, PE:EtOAc, 1:4) detectable under UV short wavelength, is a phenolic compound from its positive reaction (an orange spot) on TLC with 10% FeCl₃. It also indicated by the alkaline shift from 270 nm (π - π ^{*} transition) to 315 nm (n- π ^{*} transition) of UV absorption bands (Figure 6 and Table 3).The assignment of IR absorption bands (Figure 5) is summarized in Table (2). From the close resemblance between the spectra of <u>M</u>₅ and chlorogenic acid (Figure 5), <u>M</u>₅ should be chlorogenic acid. Chlorogenic acid is also previously reported in the plant (Stuardxchange, 2016).

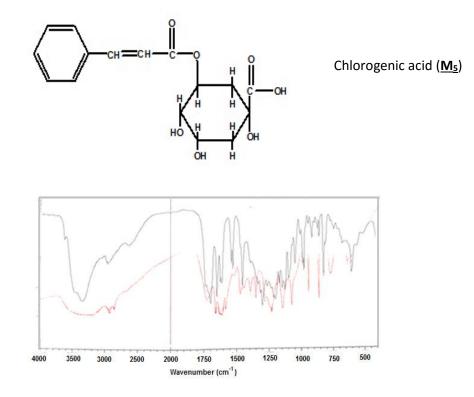


Figure 5 Overlaid IR spectra of M₅ (red) and chlorogenic acid (black) (AIST)

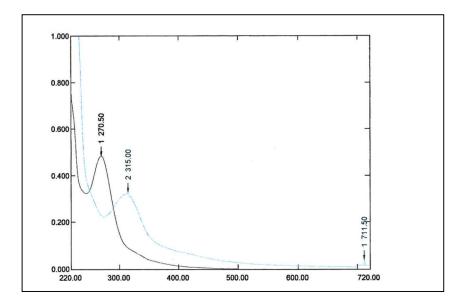


Figure 6 UV spectra of isolated compound M₅ (chlorogenic acid) from yacon tuber

The isolated compound \underline{M}_6 (R_f 0.34, Silica gel GF₂₅₄, EtOAc) was also invisible under UV short and long wavelengths on TLC plate, but could be visualized as violet spot upon heating with anisaldehyde-sulphuric acid reagent, suggesting a steroid/terpenoid compound. But its higher polarity suggested by its TLC solvent system and intense IR absorption bands for O-H and C-O groups (Figure 7) indicate a glycoside. Apart from these two absorption bands, the remaining bands are very similar to β -sitosterol. Furthermore, the observed bands are very similar to those reported for β sitosterol β -D-glucoside. Assignment of bands for <u>M</u>₆ compared with the reported bands (Arunachalam *et al.*, 2009) for β -sitosterol β -D-glucoside is given in Tables 2 and 3.Therefore <u>M</u>₆ should be β -sitosterol β -D-glucoside.

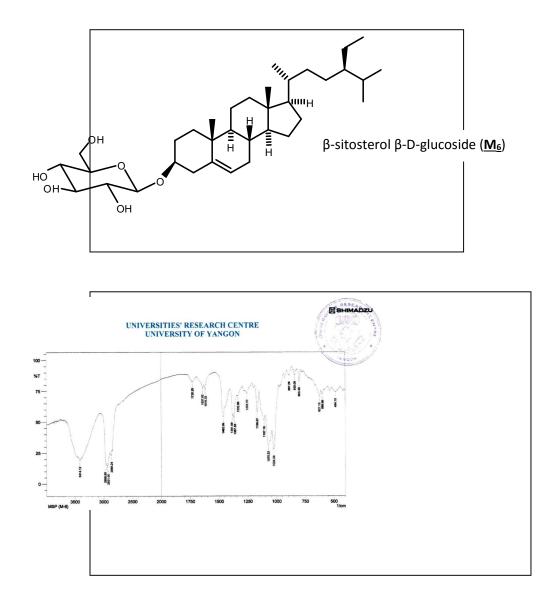


Figure 7 FTIR spectrum of isolated compound M_6 (β -sitosterol β -D-glucoside) from yacon tubers

| | Wavenumber (cm ⁻¹) | | | | | | | | | |
|----------------|--------------------------------|-------|----------------|---|---------------|---------------------------------|---------|---------------|--------------------------------|-------------|
| Sample code | V O-H | v=C-H | v C-H | First overto ne of δ CH ₂ | v C=O | V C=C | δas C-H | δs C-H | v C-O | δоор=С Н |
| M_1 | - | - | 2957, | - | 1739 | - | 1464 | 1383 | 1261 | 804, |
| | | | 2926, 2855 | | | | | | | 723, 702 |
| M ₂ | - | - | 2960, 2926, | - | 1738 | - | 1464 | 1379 | 1287 | 800 |
| M ₃ | 3550- | - | 2855 2970, | 2675 | 1697 | - | 1466, | 1350 | 1296, | 939, |
| | 2500 | | 2840 | | | | 1412 | | 1229, 1207 | 723 |
| M_4 | 3447 | - | 2980, 2850 | - | - | - | 1464 | 1381, 1370 | 1064, 1074 | 839, 800 |
| M5 | 3550- 2500 | 3100 | 2970, 2850 | - | 1726, 1659 | 1613, 1580, 1520, 1470 | 1470 | - | 1144, 1074 | 943, 864 |
| M ₆ | 3414 | - | 2960, 2850 | - | - | - | 1462 | 1381,1 368 | 1167, 1107 1070, 1024 | 839, 800 |

 Table 2
 FTIR Spectral Data of The Isolated Compounds from Ethyl Acetate Extraction of Yacon Tuber

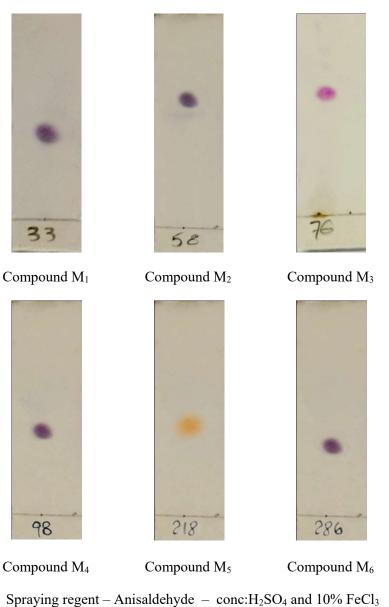
Table 3UV Spectral Data Assignment of Compound M5

| Reagents | Observed $\lambda_{max}(nm)$ | Assignment |
|----------------------|------------------------------|------------------------------------|
| Sample + MeOH | 270.50 | $\pi \rightarrow \pi^*$ transition |
| Sample + MeOH + NaOH | 315.00 | $n \rightarrow \pi^*$ transition |

| Wavenum | ıber (cm ⁻¹) | | Group assignment | |
|---------------------------|--------------------------------|-----------------------------------|--|--|
| Isolated Compound | β-sitosterol β-D-glucoside* | - Vibrational mode | | |
| 3414 | 3395 | ν О-Н | -ОН | |
| 2960-2850 | 2934, 2870 | ν С-Н | -CH ₃ ,)CH ₂ , ⇒ CH | |
| 1462 | 1461 | $\delta CH_2 \& \delta_{as} CH_3$ | >CH ₂ , -CH ₃ | |
| 1381, 1368 | 1373 | $\delta_{sy}CH_3$ | - CH ₃ | |
| 1167, 1107, 1070, 1024 | 1072, 1024 | ν C-O | 2° alcohol & ether | |
| 839, 800 | - | $\delta_{oop} = C\text{-}H$ | trisubstituted double bond | |

Table 4Comparison of FT IR Spectral Data of Compound M_6 and β -sitosterol
 β -D-glucoside

*Arunachalam et al., 2009



Solvent system – PE : EA – 99:1, 99:1, 9:1, 5:1, 1:4, EA only R_f value of M_1 , M_2 , M_3 , M_4 , M_5 , M_6 – 0.43, 0.63, 0.65, 0.44, 0.42, 0.34

Figure 8 Thin layer chromatograms of six isolated compounds from EA extract of yacon tuber

Conclusion

The results of the present study established the presence of phytochemicals in the yacon tuber extract. Qualitative phytochemical screening was carried out to establish the presence of alkaloids, flavonoids, polyphenols, α -amino acids, glycosides, steroids, terpenoids, phenolic compounds, carbohydrate, saponins, tannins, protein and reducing sugars in the yacon tuber. Starch and cyanogenic glycosides were not detected. The experimental data also suggested the presence of significant amounts of stigmasteryl acetate (0.003%), β -sitosteryl acetate (0.005%), hexadecanoic acid (0.009%), β -sitosterol (0.02%), chlorogenic acid (0.12%) and β -sitosterol β -D-glucoside (0.8%) in the ethyl acetate fraction of the 70% ethanol extract of the yacon tuber. Thus yacon, *i.e.Smallanthus sonchifolius* tubers has great potential for the human health.

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